

Linkage of Bioinformatics, Computer Sciences, and Experimental Sciences
Prince William Campus, Manassas, VA

## **Environmental Biocomplexity Monitoring Microbial Communities**

#### **Objectives**

#### Patrick Gillevet

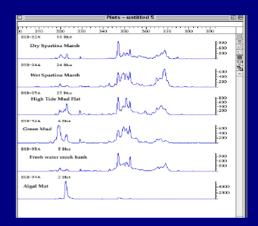
- Comprehensive continuous field monitoring architecture for bacterial, protists, and fungi
- Understand behavior of groups of organisms and their response to environmental change

#### Approach

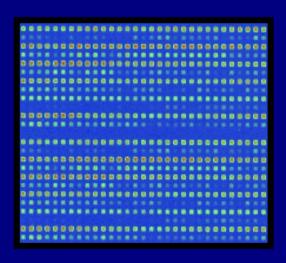
- Amplicon Length Heterogeneity (ALH)
  - Continuous monitoring of community dynamics
  - SSU rRNA, ITS, functional genes
- Sequencing SSU rRNA clone libraries
  - Initial characterization of community
- Microarray technology
  - Rapid characterization of samples of interest (verification)

#### **Applications**

- Risk assessment of genetically modified organisms
- Monitoring polymicrobial diseases -Crohn's disease
- Monitoring bioremediation processes
- Complete Background Characterization
- Monitoring Biowarfare agents & interferants



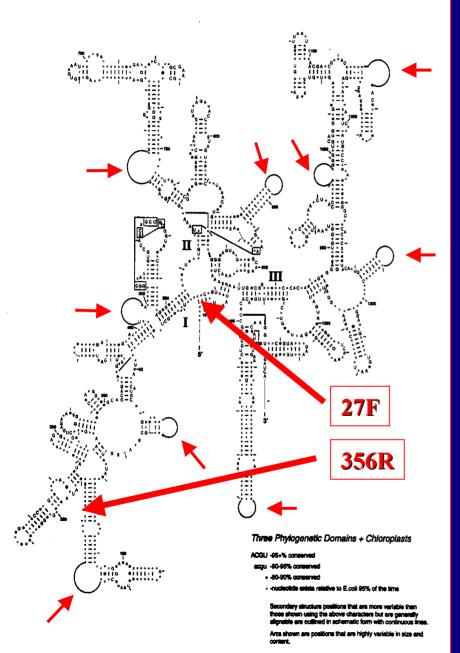
#### **ALH Fingerprinting**



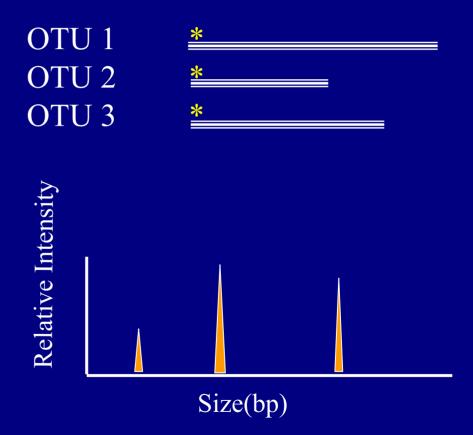
Oligo Microarrays

Sponsors: NSF, Sea Grant, CBNP

Phylogenetic conservation superimposed onto the Escherichia coli small subunit ribosomal RNA secondary structure



#### Amplicon Length Heterogeneity Fingerprinting (ALH)

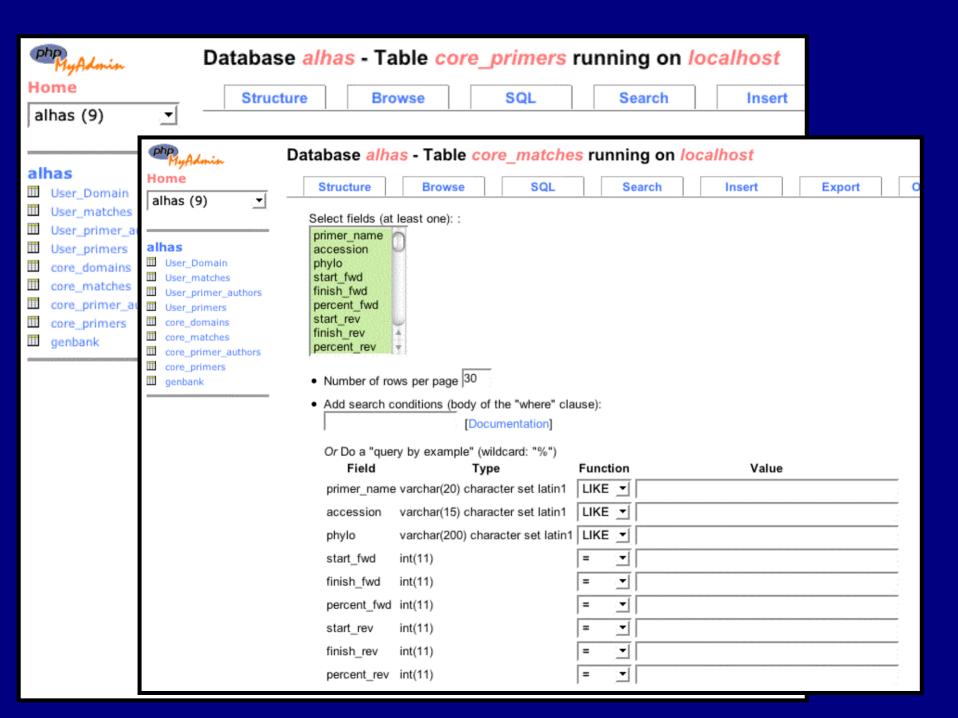


Peak area ~ Abundance

# Ampleion Length Heretogeneity Analysis System (ALHAS)

- P. Gillevet, J. Grefenstette, L. Kumar, A. Ahmed, A. Rehman
  - Compile rRNA data from Genbank
  - Calculate amplicon length (Domain) from primers
  - Determine specificity of Domain
  - Predict community composition from fingerprint

	Primer Submission Page							
First Name:								
Last Name:								
Email:								
Primer I:								
Primer I Specificity:								
Primer I Sequence:								
Primer II:								
Primer II Specificity:								
Primer II Sequence:								
Reference:								
	Type in your comments here							



### Ecoinformatics Tools for Microbial Diversity Studies: Supervised Classification of Amplicon Length Heterogeneity (ALH) Profiles of 16S rRNA

Chengyong Yang, Yong Wang, DeEtta Mills, Kalai Mathee, Krish Jayachandran, Masoumeh Sikaroodi, Patrick Gillevet, Jim Entry, Giri Narasimhan George Mason University and Florida International University

#### Supervised classification tools:

- (a) Support Vector Machines (SVM)
- (b) K-Nearest Neighbor Method (KNN)

Analyzed 4 amplicons: V1+V2 6-FAM-27F and 355R

V1 6-FAM-P1F and P1R

V3 HEX-338F and 518R

V9 NED-1055F and EC1392R

SVM was able to classify Idaho soils but not Chesapeake Bay sediments V1+V2 was most informative

## **Automatically Megablast against RDP database Sort by RDP Number**

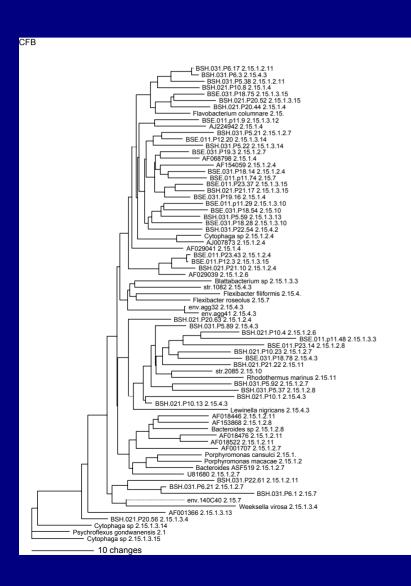
BSE A BSH A BSE B BSH B Total Level 3/4 Total LEVEL 1

RDP number

Description

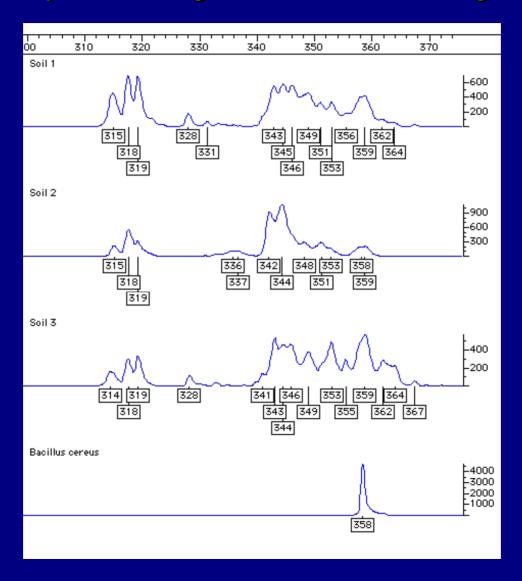
KDP Hulliber	Description	DOL_M DO	A	DOC_D	DOU D	IUCAI	Level 3/4 local		
1.2.2.1.4	ENVIRONMENTAL_CLONE_4B7_SUBGROUP				1	1	1	ARCHAEA	CRENARCHAEOTA
2.1	THERMOPHILIC_OXYGEN_REDUCERS				1	1		BACTEDIA	THERMOPHILIC_OXYGEN_REDUCERS
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2.3	CTM.PROTEOLYTICUS_GROUP				1	1	2	BACTERIA	CTM.PROTEOLYTICUS_GROUP
2.7.1.2	ENVIRONMENTAL_CLONE_T78_GROUP		1		1	2		BACTERIA	GREEN_NON-SULFUR_BACTERIA_AND_RELATIVES
2.7.2.1.3	MEI.RUBER_SUBGROUP	1		100		1	3		GREEN_NON-SULFUR_BACTERIA_AND_RELATIVES
2.9.1.2	NSP.MOSCOVIENSIS_SUBGROUP				1	1		BACTEDIA	LEPTOSPIRILLUM-NITROSPIRA
2.9.3.1	LPP.FERROOXIDANS_SUBGROUP				1	1			LEPTOSPIRILLUM-NITROSPIRA
2.9.4	TDV.YELLOWSTONII_GROUP	1		1		2	4		LEPTOSPIRILLUM-NITROSPIRA
2.7.4	TDV.YELLOWSTONII_GROUP	1				-		BACTERIA	LEPTOSPIRILLOM-NITROSPIRA
2.10.1	ENVIRONMENTAL_CLONE_WCHB1-41_SUBGROUP			1		1		BACTERIA	PROSTHECOBACTER_GROUP
2.10.2	VER.SP_STR_VEGLC2_SUBGROUP			1		1	2	BACTERIA	PROSTHECOBACTER_GROUP
2.12	ENVIRONMENTAL_CLONE_OPB2_GROUP	1		1	1	3	3	BACTERIA	ENVIRONMENTAL_CLONE_OPB2_GROUP
2.12	ENVIRONMENTAL_CLONE_OPB2_GROUP	•				•	•	BACTERIA	ENVIRONMENTAL_CLONE_OPB2_GROUP
2.13.1	ENVIRONMENTAL_CLONE_JAP604_GROUP		1			1			NITROSPINA_SUBDIVISION
2.13.4	ENVIRONMENTAL_CLONE_OPB5_GROUP		1		1	2		BACTERIA	NITROSPINA_SUBDIVISION
2.13.5	ENVIRONMENTAL_CLONE_RB25_GROUP				1	1		BACTERIA	NITROSPINA_SUBDIVISION
2.13.6	ENVIRONMENTAL_CLONE_III1-8_GROUP			1		1			NITROSPINA_SUBDIVISION
2.13.8	HP.FOETIDA_GROUP				1	1		BACTERIA	NITROSPINA_SUBDIVISION
2.13.12	ENVIRONMENTAL_CLONE_RB40_GROUP		1			1	7		NITROSPINA_SUBDIVISION
2.14	FLS.SINUSARABICI_ASSEMBLAGE		1			1	1	BACTERIA	FLS.SINUSARABICI_ASSEMBLAGE
2.15.1.2.4	CY.FERMENTANS_SUBGROUP	1	2	1		4		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.2.6	BAC,SPLANCHNICUS SUBGROUP		1			1			FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.2.7	PPM.MACACAE_SUBGROUP		1	1	3	5			FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.2.8	BAC.FRAGILIS_SUBGROUP	1			1	2			FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.2.11	PRV.RUMINICOLA_SUBGROUP				3	3	15		FLEXIBACTER-CYTOPHAGA-BACTEROIDES
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2.15.1.3.3	BLT.SP_SUBGROUP	1				1		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.3.4	EMB.BREVIS_SUBGROUP		1			1		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.3.10	F.FLEVENSE_SUBGROUP	1		1		2			FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.3.12	CAP.OCHRACEA_SUBGROUP	1				1			FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.3.13	CY.ULIGINOSA_SUBGROUP				1	1		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.3.14	PSF.TORQUIS_SUBGROUP	1			1	2		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.3.15	PSH.BURTONENSIS_SUBGROUP	2	2	1		5		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.1.4	CYTOPHAGA_GROUP_II		2	1		3	16	BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.4.2	FLX.SANCTI_SUBGROUP				1	1		BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.4.3	LEW.NIGRICANS_SUBGROUP		2	1	2	5	6		FLEXIBACTER-CYTOPHAGA-BACTEROIDES
21101410	EERINIONIO GODONO OF				-			DHOTEKIM	TEENIDISTER OTTOFTINGH BHOTEROIDES
2.15.7	FLX.LITORALIS_GROUP	1			1	2	2	BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.8	TMN.LAPSUM_GROUP				1	1	1	BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.15.10	STR.SBR2085			1		1	1	BACTERIA	FLEXIBACTER-CYTOPHAGA-BACTEROIDES
2.10/10							•	Z. TO TEXT	Snapz Pro X
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## **Automatic Report Generation**





### Survey of Backgrounds in Washington DC

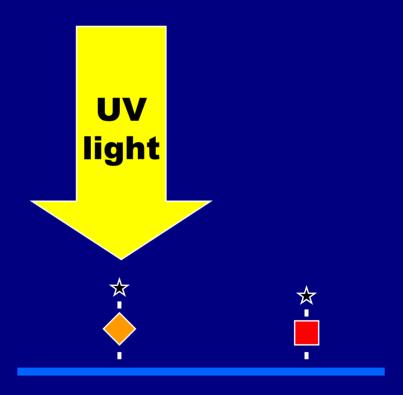




# Oligo Synthesis Controlled by Light 2.5 orders of magnitude reduction in cost

Steve Smith

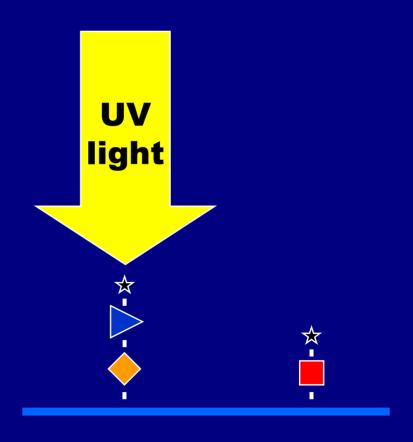
- ☆ Photo labile protecting group
- DNA monomer





## Oligo Synthesis Controlled by Light

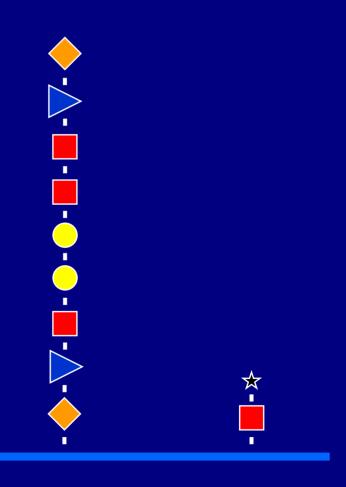
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- DNA monomer





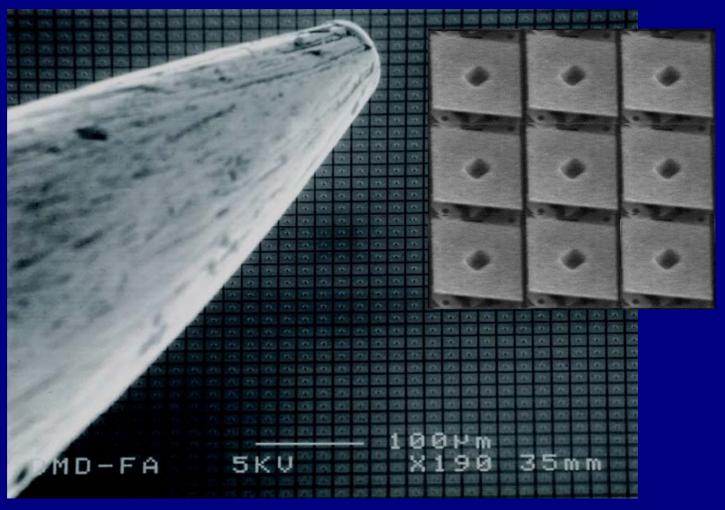
## Oligo Synthesis Controlled by Light

- ☆ Photo labile protecting group
- DNA monomer





#### DMD: Digital Micromirror Device

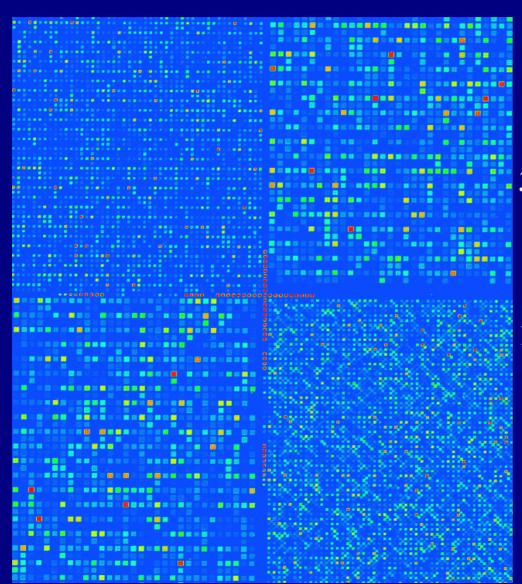


195,000 (37mer - 70 mer)

390,000 (< 36mer)



## **Array Options**

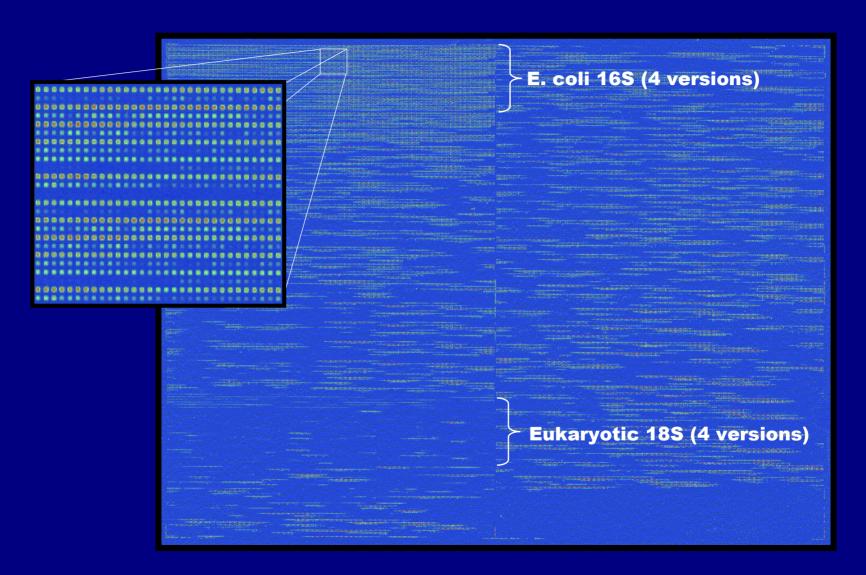


■ 1 - 36mers
390,000 features per array

**37 - 70mers 195,000 features per array** 



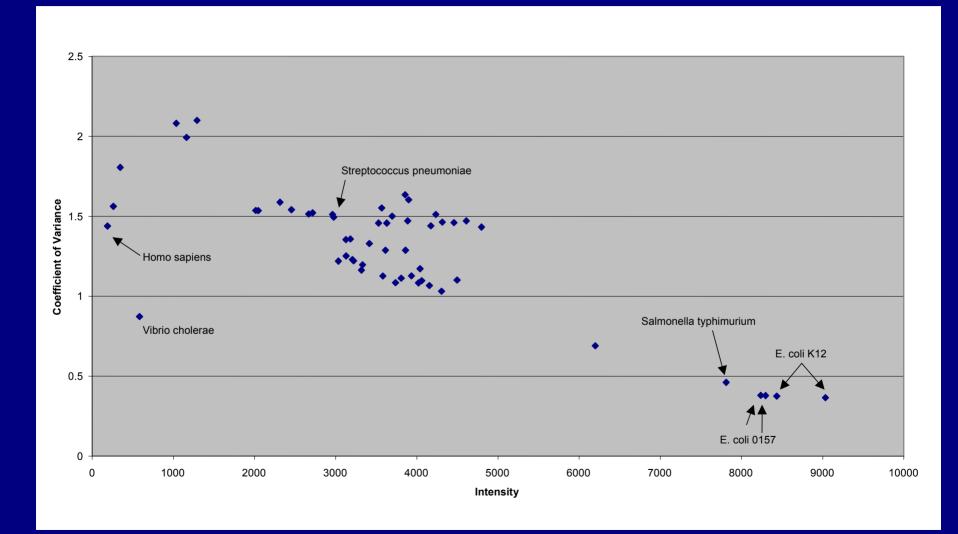
## Tiling Ribosomal RNA of 53 Species



Hybridized to E. coli K12 RNA

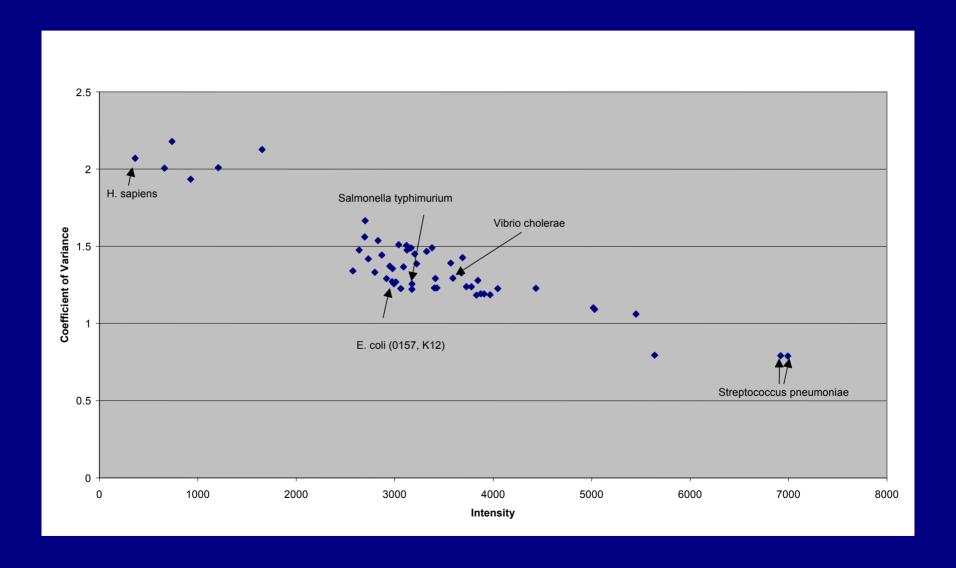


### 79 oligos x 55 Taxa Array Hybridized to E. coli K12 RNA



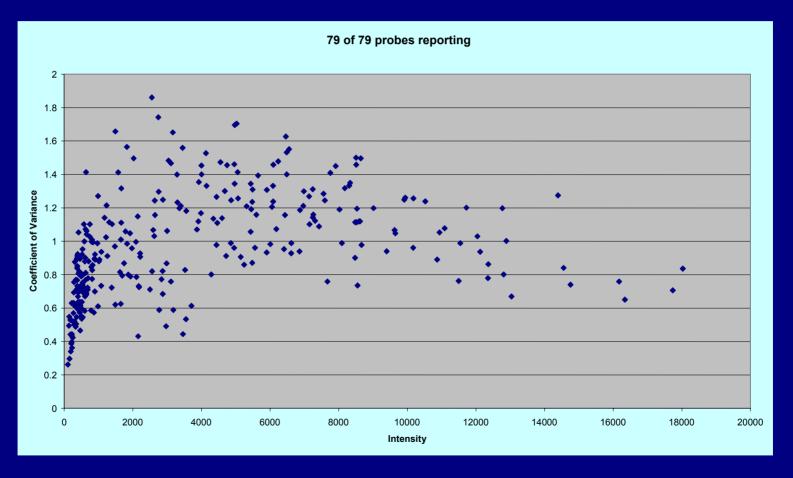


### 79 oligos to 55 Taxa Array Hybridized to Oral Swab DNA





### 79 probes to 4816 Taxa Array 390,000 oligos Hybridized to Oral Swab DNA



GMU: Possible use of supervised and unsupervised methods
Combine ALH and Microarray data to deconvolve community

#### **Bioinformatics Solutions**

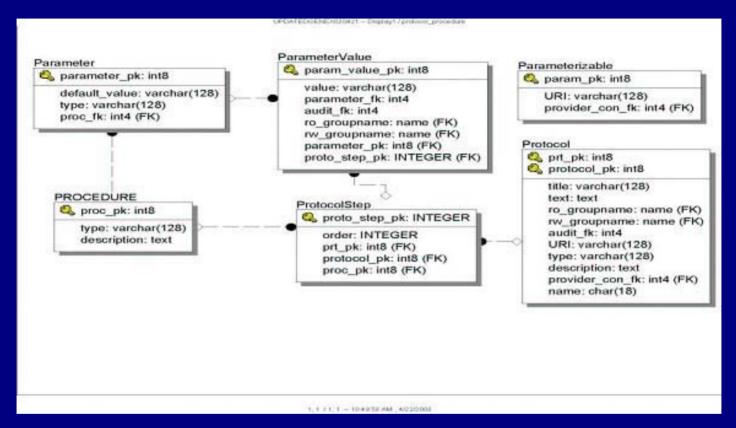
Jennifer W. Weller

• Problem: Data from disparate sources are combined in one analysis. How do you track the source and the evidence- weight chosen in a particular analysis?

#### Solutions

- LIMS systems and e-notebooks (in concert with bar coding) track the processes of acquiring physical data.
- Database handling of branched protocols (with method- or investigator- specific parameters) that can be automatically called.

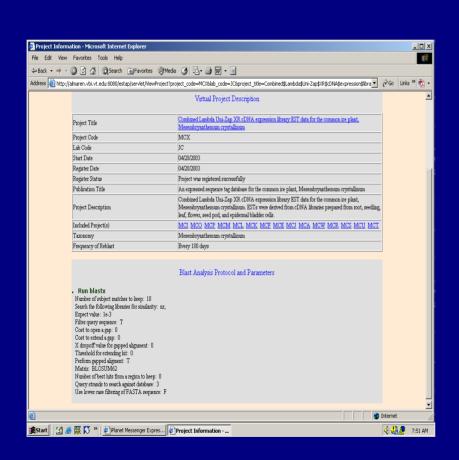
#### **BioInformatics Database Solutions**

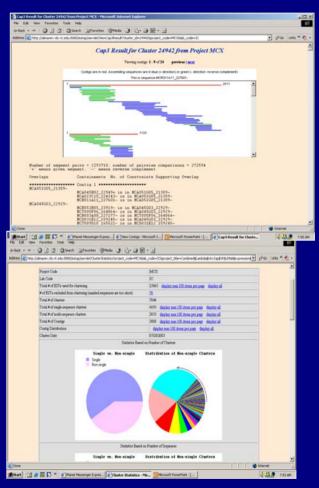


- The goal: track how data are transformed
- The basic components are reusable and reconfigurable
  - currently implemented using perl scripts for PostgreSQL

## **Current Implementations**

• ESTAP : EST analysis pipeline



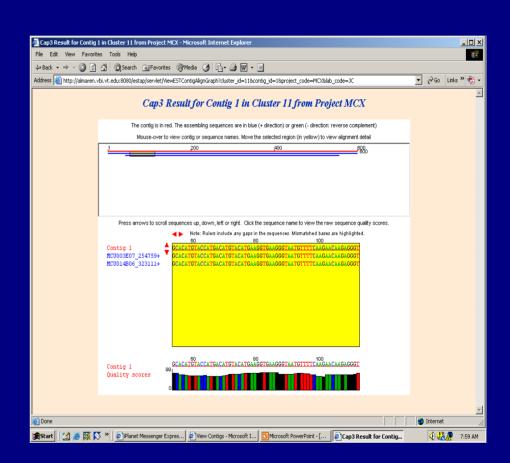


#### ESTAP continued

There are many ways to cluster ESTs.

SNP discovery comes from clustering – for some applications the quality score may be more rigorously controlled than for others.

The underlying data model allows parallel analyses to be run and stored, then called for a particular application.



## GeneX Microarray Database Jennifer Weller

#### Objectives

- Organize large microarray datasets for sharing, analysis and data mining
- Develop analysis methods that provide insight into biological processes based on microarray data

#### Approach

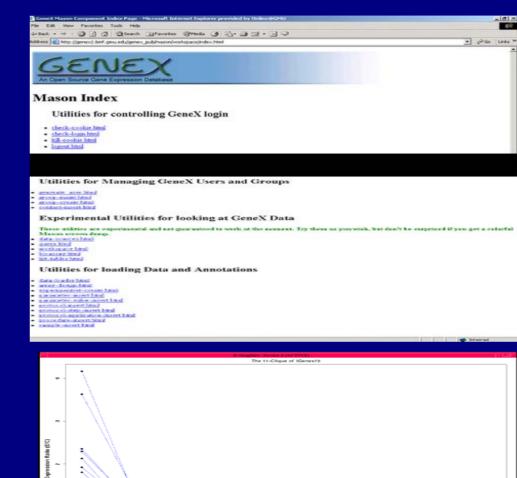
- Develop a reference implementation of the MAGE model (GeneX)
- Populate the GeneX database to test and refine performance
- Develop APIs to the database for ease in testing new analysis methods

#### Collaborators

CalTech, UVa, VCU

#### Sponsors (Current)

NSF

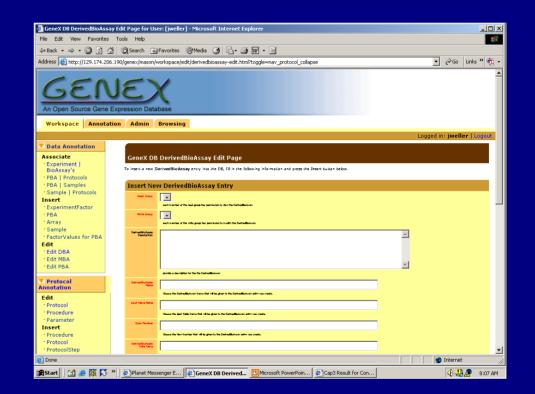


## Workspace Concept

GeneX has Project- and Investigator-specific Views, that we call Workspaces.

It is possible to define new Tables for the Workspace, allowing greater flexibility for storing the results of computational experiments.

We are working on providing code that will generate basic Reports from these new tables automatically.



## Academic Programs at GMU www.gmu.edu

- Bioinformatics and Computational Biology
  - Ph.D. in Bioinformatics (1992)
  - M.S. in Bioinformatics (2002)
- www.binf.gmu.edu
- Computational Science and Informatics
  - Ph.D. in Computational Neurobiology
  - M.S. in CSI
  - Certificate in CSI
- Environmental Sciences and BioSciences (CAS)
  - Ph.D in Environmental Sciences and Policy
  - Ph.D. in BioSciences
  - M.S. in Biology
- mason.gmu.edu/~esp

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